Novel endophytic nematode antagonistic fungi – potential for nematode biocontrol

Samad Ashrafi¹, Abdelfattah A. Dababat², Wolfgang Maier¹

¹Institute for Epidemiology and Pathogen Diagnostics, Julius Kühn-Institut (JKI)—Federal Research Centre for Cultivated Plants, Braunschweig, Germany ²International Maize and Wheat Improvement Center (CIMMYT), Ankara, Turkey



Overview

Cereal cyst nematodes (CCNs) can lead to significant yield reductions of grains. The use of nematicidal chemicals is banned in many countries due to their generally high non-target toxicity. Antagonistic microorganisms controlling nematodes are an important alternative.

Microscopic observations of cyst samples of the CCN *Heterodera filipjevi* obtained from wheat fields in Turkey regularly revealed nematode cysts containing fungal colonised eggs (Fig. 1). The aim of this study was to:

Results

The approach applied in this study resulted in finding of eight new fungal species, all isolated from symptomatic nematode eggs. The newly described *ljuhya vitellina* and *Monocillium gamsii* belong to the Bionectriaceae and Niessliaceae, respectively. The new fungal genus *Polyphilus* was proposed to accommodate the new species *P. sieberi* representing a dark septate endophyte (DSE). Two more species were preliminarily characterized as DSEs belonging to the Pleosporales. These are the first DSEs found to parasitize nematode eggs. All newly-found species could be successfully re-isolated from artificially infected nematodes and Koch's postulates were thus fulfilled (Fig. 3A-C).

- isolate and classify fungi isolated from nematode eggs, and fulfil Koch's postulates,
- study the nematode-fungus interaction microscopically,
- identify secondary metabolites produced by these fungi.



Fig. 1. Symptomatic eggs of *H. filipjevi* colonised with different fungal species. Several novel fungal species producing new natural compounds were found during this study.

Methodology

A single-egg based technique was developed to isolate fungi from



Fig. 3A. *Ijuhya vitellina.* A, B) Cysts and eggs of *H. filipjevi* naturally infested with *I. vitellina.* Infested eggs accommodate one, occasionally two globose to subglobose microsclerotia. C) Transformation of hyphae into microsclerotium. D) *In vitro* colonisation of a nematode egg by hyphae developed into microsclerotium [1].



Fig. 3B. Monocillium gamsii. A, B) Naturally infested cysts and eggs of H. filipjevi naturally infested with M. gamsii. Infested eggs contain one or two microsclerotia. C) phialides and conidia. D) Formation of microsclerotia by development of dictyochlamydospore structures [2].



symptomatic nematode eggs (Fig. 2A):

- Collection of diseased eggs from each symptomatic cyst
- Individually plating of single eggs on agar plates
- Fungal identification and taxonomy
- In vitro examination of isolated fungi against nematode eggs

Metabolite profiling of the discovered fungi (Fig. 2B):

- Small and medium-scale fermentation of fungal cultures
- Application of EtOAc and HPLC-based techniques for extraction, purification and structural elucidation of secondary metabolites
- Nematode bioassays and cytotoxicity of obtained compounds



Fig. 3C. Polyphilus sieberi. A, B) Cysts and eggs of H. filipjevi naturally infested with P. sieberi. Infested eggs were colonised with moniliform and strongly melanised hyphae. C, D) fungal colonisation of a nematode juvenile and nematode egg in vitro [3].

Four novel compounds with nematicidal activity were isolated from a hitherto undescribed pleosporalean strain (DSM106825) belonging to the family of Phaeosphaeriaceae (Fig. 4A-E) that parasitises eggs of *H. filipjevi* [4].



Fig. 2. A general overview of the methods applied in this study. A) methodes applied in cultural studies. B) methods applied for secondary metabolite profiling

Arthrichitin (3): $R_1 = =0$; $R_2 = H$ Arthrichitin B (4): $R_1 = H_2$; $R_2 = H$ Arthrichitin C (5): $R_1 = =0$; $R_2 = CH_3$

Xanthomide Z

Fig. 4. A new Phaeosphaeriaceous fungus isolated from eggs of *H. filipjevi*, and the source of the here presented new compounds. **A)** BI of the strain DSM106825 based on mutil-gene (ITS-LSU-TEF1) phylogenetic analyses. **B)** A pure culture of the fungus. **C)** The new compounds isolated from the fungus.

References

- 1. Ashrafi, S., S. Helaly, H.-J. Schroers, M. Stadler, K. R. Richert-Poeggeler, A. A. Dababat and W. Maier (2017a). *Ijuhya vitellina* sp. nov., a novel source for chaetoglobosin A, is a destructive parasite of the cereal cyst nematode *Heterodera filipjevi*. PLOS ONE 12(7)
- 2. Ashrafi, S., M. Stadler, A. A. Dababat, K. R. Richert-Pöggeler, M. R. Finckh and W. Maier (2017b). *Monocillium gamsii* sp. nov. and *Monocillium bulbillosum*: two nematode-associated fungi parasitising the eggs of *Heterodera filipjevi*. Mycokeys 27: 21-38
- Ashrafi, S., D. G. Knapp, D. Blaudez, M. Chalot, J. G. Maciá-Vicente, I. Zagyva, A. A. Dababat, W. Maier and G. M. Kovács (2018). Inhabiting plant roots, nematodes and truffles — Polyphilus, a new helotialean genus with two globally distributed species. Mycologia 110(2): 286-299.
- 4. Helaly S. E., S. Ashrafi, R. Teponno, S. Bernecker, A. A. Dababat, W. Maier, and M. Stadler (2018). Nematicidal cyclic lipodepsipeptides and a xanthocillin derivative, from a pleosporalean fungus parasitizing eggs of the plant parasitic nematode *Heterodera filipjevi*. Journal of Natural Products (Accepted)

E-Mail: samad.ashrafi@julius-kuehn.de

Institute for Epidemiology and Pathogen Diagnostics, Messeweg 11-12, 38104 Braunschweig



www.julius-kuehn.de